

SYNTHESIS OF 24,24-DIFLUORO-25-HYDROXYVITAMIN D₃

Sachiko Yamada, Masayuki Ohmori, and Hiroaki Takayama*
Faculty of Pharmaceutical Sciences, Teikyo University
Sagamiko, Kanagawa 199-01 Japan

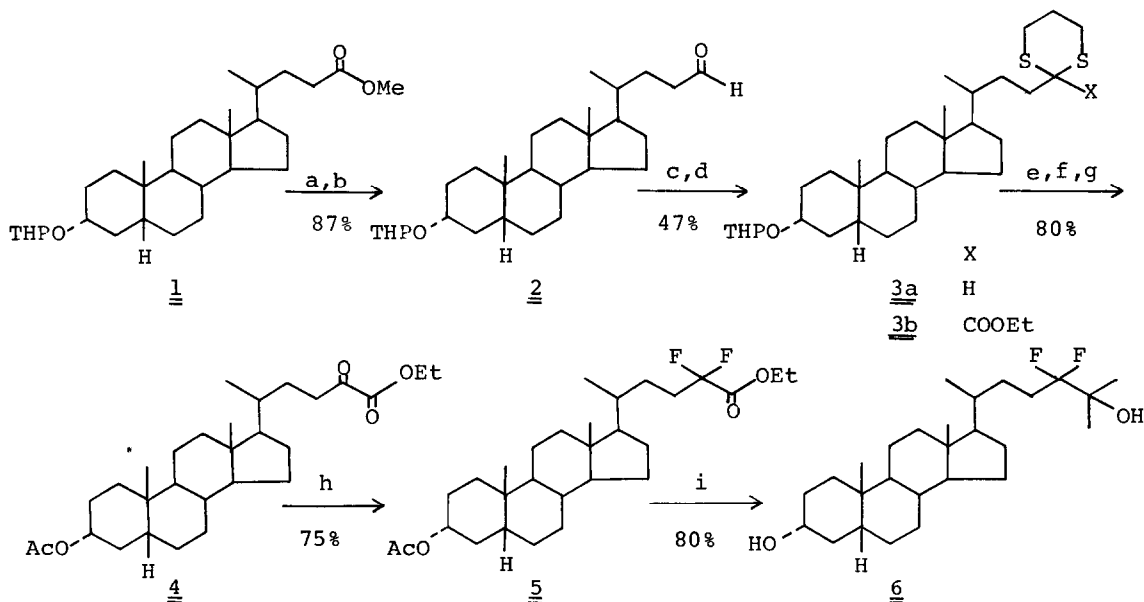
Summary: 24,24-Difluoro-25-hydroxyvitamin D₃ (13) has been prepared from commercially available lithocholic acid derivative to study the role of 24-hydroxylation in the metabolism of vitamin D₃.

By extensive studies on the metabolism of vitamin D₃, a number of metabolites have been isolated and identified.¹ Besides the active metabolite (i.e. 1 α ,25-dihydroxyvitamin D₃) and the precursor (i.e. 25-hydroxyvitamin D₃) leading to it, the metabolites hydroxylated at 24-position [i.e. (24R)-24,25-dihydroxyvitamin D₃ and (24R)-1 α ,24,25-trihydroxyvitamin D₃] have been directed much attention because of their reduced but still considerably high activity compared with the corresponding non-hydroxylated vitamin.² However, the role of 24-hydroxylated vitamin D in the metabolism of the vitamin has still been remained to be clarified.

Due to similar steric bulk and dissimilar chemical behavior, fluorinated biological compounds have been known to act as antimetabolites with respect to the corresponding fluorine-free natural products.³ Based on this reason, we have chosen the title compound (13) in which 24-position is blocked for metabolic hydroxylation by substitution with fluorine atoms as an analog of 25-hydroxyvitamin D₃ to study the role of 24-hydroxylation in the metabolism of the vitamin.

Using commercially available lithocholic acid, construction of the side chain was achieved as shown in Scheme 1 in which the desired α,α -difluoro- \bar{t} -carbinol function was derived efficiently from the α -keto ester, and the key intermediate, 24,24-difluoro-5 β -cholestane-3 α ,25-diol (6), was obtained in 20% overall yield. Reduction of the ester 1 with LiAlH₄ followed by Collins oxidation gave the aldehyde 2: IR (CHCl₃) 1715 cm⁻¹; NMR (CDCl₃) δ 0.65 (3H, s), 0.97 (3H, s), 4.75 (1H, m), 9.82 (1H, t, J = 2 Hz). The 1,3-dithiane 3a derived from the aldehyde 2 was transformed into the ester 3b [IR (CHCl₃) 1720 cm⁻¹; NMR (CDCl₃) δ 0.63 (3H, s), 0.91 (3H, s), 1.34 (3H, t, J = 6.5 Hz), 4.27 (2H, q, J = 6.5 Hz), 4.74 (1H, m)] via its lithium salt by reaction with ethyl chloroformate.⁴ The α -keto ester 4 [IR (CHCl₃) 1720 cm⁻¹; m/e 474, 414] obtained by exchanging the protecting group of the alcohol from THP to acetyl followed by removal of the dithiane group⁵ was treated with diethylaminosulfur trifluoride (DAST)⁶ to provide the difluoro ester 5 [IR (CHCl₃) 1755, 1720 cm⁻¹; NMR (CDCl₃) δ 0.65 (3H, s), 0.93 (3H, s), 1.36 (3H, t, J =

7 Hz), 2.03 (3H, s), 4.35 (2H, q, $J = 7$ Hz), 4.73 (1H, m)], which upon treatment with methyl magnesium bromide afforded the α -carbinol 6: 142-143°; NMR (CDCl_3) δ 0.66 (3H, s), 0.92 (3H, s), 1.31 (6H, s), 3.65 (1H, m); m/e 440, 422.



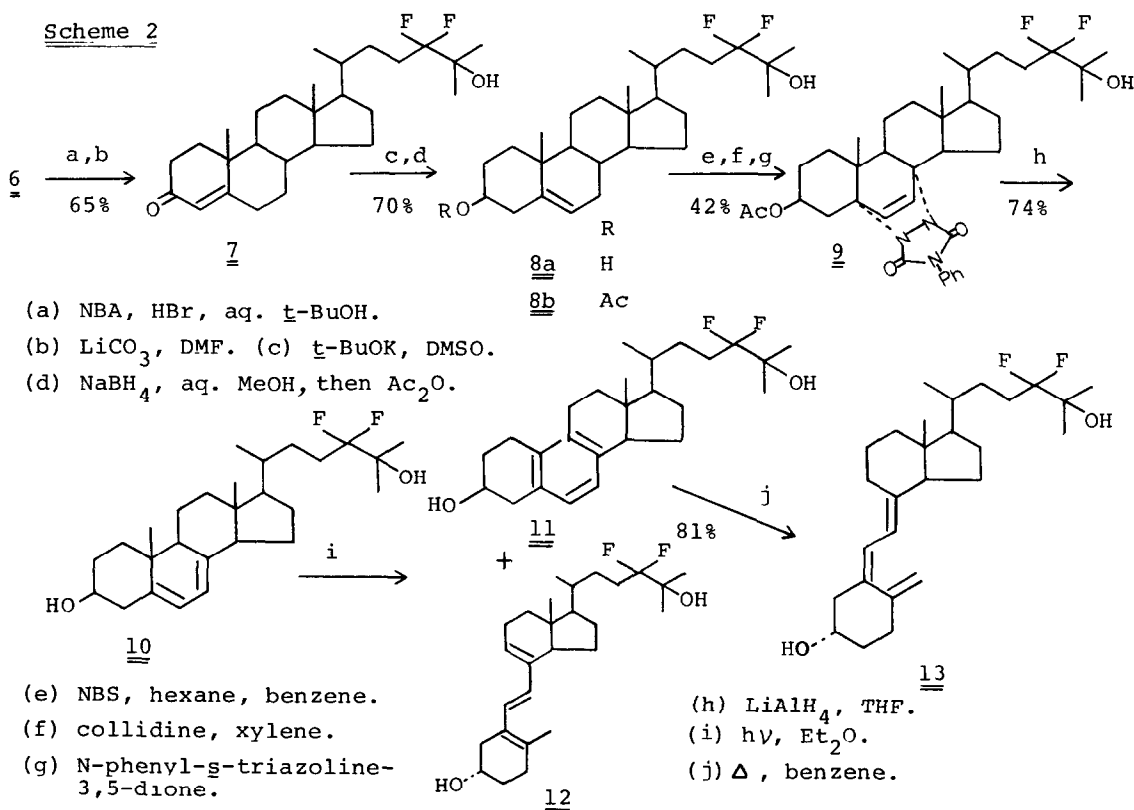
- (a) LiAlH_4 , THF. (b) CrO_3 .pyridine.HCl, CH_2Cl_2 . (c) 1,3-propanedithiol, $\text{BF}_3 \cdot \text{Et}_2\text{O}$, CH_2Cl_2 , then dihydropyran, p -TsOH.pyridine. (d) n -BuLi, THF, then ClCOEt . (e) p -TsOH.pyridine, EtOH. (f) Ac_2O , pyridine. (g) NBS, aq. acetone. (h) DAST, CH_2Cl_2 . (i) CH_3MgBr , THF.

Scheme 1

Having thus established the construction of the side chain, the stage was set to effect the transformation of the saturated steroid skeleton to the vitamin D ring system. Conversion to the 3β -hydroxy- Δ^5 -steroid 8a [mp 168-170°; m/e 438, 420; NMR (CDCl_3) δ 0.70 (3H, s), 0.95 (3H, d, $J = 6$ Hz), 1.02 (3H, s), 1.32 (6H, s), 3.55 (1H, m, $w/2 = 24$ Hz), 5.40 (1H, d, $J = 4$ Hz)] was achieved by deconjugation⁷ of the enone 7 (mp 177-178°; m/e 436), which was obtained by simultaneous oxidation bromination reaction⁸ of 6 followed by dehydrobromination, and subsequent reduction with NaBH_4 . The acetate 8b derived from 8a was transformed to the corresponding vitamin in the usual manner.⁹ Allylic bromination followed by dehydrobromination gave a mixture of the 5,7- and the 4,6-diene from which the former was isolated as the triazoline adduct 9 mp 194-195°; NMR (CDCl_3) δ 0.82 (3H, s), 1.00 (3H, s), 1.32 (6H, s), 2.04 (3H, s), 3.25 (1H, dd, $J = 14, 5$ Hz), 5.50 (1H, tt, $J = 10, 5$ Hz), 6.36 (2H, ABq, $J = 8$ Hz), which by reduction with LiAlH_4 afforded the provitamin 10: mp 177-179°; m/e 436, 403, 377; NMR (CDCl_3) δ 0.63 (3H, s), 0.94 (3H, s), 1.31 (6H, s), 3.60 (1H, m, $w/2 = 26$ Hz), 5.40 (1H, m), 5.60 (1H, dd, $J = 6, 2$ Hz). Irradiation

of the provitamin 10 in ether by high pressure mercury lamp through Vycor filter and chromatography of the products on Sephadex LH20 yielded the pre-vitamin 11 (38.5%) [λ_{\max} (95% EtOH) 262 nm], the tachysterol analog¹⁰ 12 (25.5%) [λ_{\max} (95% EtOH) 271(sh), 281, 291 (sh) nm; NMR (CDCl₃) δ 0.72 (3H, s), 1.00 (3H, d, $J = 6$ Hz), 1.34 (6H, s), 1.82 (3H, s), 4.00 (1H, m), 5.75 (1H, bs) 6.40 (2H, ABq, $J = 16$ Hz)], and unchanged 10 (7.5%). Refluxing in benzene (2 hr) and subsequent storage at room temperature (5 days) converted the pre-vitamin 11 exclusively to the vitamin 13 (81%). The vitamin thus obtained showed typical UV spectrum [λ_{\max} (95% EtOH) 265 nm ($\log \epsilon = 4.24$)] and mass fragmentation pattern (m/e 436, 136, 118) of vitamin D derivatives supporting the assigned structure. The ¹H NMR spectrum of 13 afforded further supporting evidence for the structure showing the signals of methyl groups at δ 0.56 (3H, s), 0.95 (3H, d, $J = 6$ Hz), and 1.32 (6H, s), the 3 α proton at δ 3.95 as multiplet with $w/2 = 20$ Hz, the C-19 olefinic protons at δ 4.85 and 5.08 as broad singlets, and the protons on C-6 and C-7 at δ 6.16 as AB quartet with $J = 11$ Hz. (Scheme 2).

The fluorovitamin obtained in the present work was now subjected to the biological testing. The results will be reported elsewhere.



Synthesis of 24,24-difluoro-1 α ,25-dihydroxyvitamin D₃ is progressing using the same synthetic intermediate 6.

References

- 1) H. F. DeLuca and H. K. Schoes, Ann. Rev. Biochem., 45, 631 (1976).
- 2) I. T. Boyle, R. W. Gray, and H. F. DeLuca, Proc. Natl. Acad. Sci. U.S.A., 68, 2131 (1971). I. T. Boyle, J. L. Omdahl, R. W. Gray, and H. F. DeLuca, J. Biol. Chem., 248, 4174 (1973). Y. Tanaka, H. F. DeLuca, N. Ikekawa, M. Morisaki, and N. Koisumi, Arch. Biochem. Biophys., 170, 620 (1975). M. F. Holick, L. A. Baxter, P. K. Schraufrogel, T. E. Tavela, and H. F. DeLuca, J. Biol. Chem., 251, 397 (1976).
- 3) M. Schlosser, Tetrahedron, 34, 3 (1978), and references cited therein.
- 4) E. J. Corey, and D. Seebach, Angew. Chem., 77, 1134 (1965). E. J. Corey and D. Seebach, Angew. Chem., 77, 1135 (1965).
- 5) E. J. Corey and B. W. Erickson, J. Org. Chem., 36, 3553 (1971).
- 6) W. J. Middleton, J. Org. Chem., 40, 574 (1975).
- 7) E. Shapiro, L. Weber, E. P. Oliveto, H. L. Herzog, R. Neri, S. Tolksdorf, M. Tanabe, and D. F. Crow, Steroids, 9, 461 (1966). C. Kaneko, S. Yamada, A. Sugimoto, Y. Eguchi, M. Ishikawa, T. Suda, M. Suzuki, S. Kakuta, and S. Sasaki, Steroids, 23, 75 (1974).
- 8) E. B. Hershberg, C. Gerold, and E. P. Oliveto, J. Am. Chem. Soc., 74, 3849 (1949).
- 9) S. Bernstein, L. J. Binovi, L. Dorfman, K. J. Sax, and Y. Subbarow, J. Org. Chem., 14, 433 (1949).
- 10) A. L. Koevoet, A. Verloop, and E. Havinga, Recl. Trav. Chim. Pays-Bas, 74, 788 (1955).

(Received in Japan 16 March 1979)